

Original Article

Association between periodontal disease and plasma levels of cholesterol and triglycerides

Asociación entre enfermedad periodontal y niveles plasmáticos de colesterol y triglicéridos

Jaramillo Adriana^a, Lafaurie Gloria Inés^b, Millán Lina Viviana^b, Ardila Carlos Martin^c, Duque Andrés^d,
Novoa Camilo^e, López Diego^f, Contreras Adolfo^a.

^aPeriodontal Medicine Group, School of Dentistry, Universidad del Valle

^bInstituto UIBO, Faculty of Dentistry, Universidad El Bosque

^cUniversidad de Antioquia, Medellín Colombia.

^dFaculty of Dentistry, Universidad CES.

^eFaculty of Dentistry, Pontificia Universidad Javeriana.

^fSchool of Dentistry, Universidad del Valle.

Jaramillo A, Lafaurie GI, Millán LV, Ardila CM, Duque A, Novoa C, López D, Contreras A. Association between periodontal disease and plasma levels of cholesterol and triglycerides. *Colomb Med.* 2013; 44 (2): 80-6.

© 2013 Universidad del Valle. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Article history:

Received 30 May 2012

Received in revised form 10 June 2012

Accepted 3 April 2013

Available online 30 June 2013

Keywords:

Periodontal disease,
dyslipidemia, HDL, LDL,
triglycerides

Palabras clave:

Enfermedad periodontal,
dislipidemia, HDL, LDL,
triglicéridos

Abstract

Objective: Untreated periodontal disease seems to cause low grade systemic inflammation and blood lipid alteration leading to increased cardiovascular disease risk. To start testing this hypothesis in Colombian patients, a multicentre study was conducted including the three main state capitals: Bogotá, Medellín and Cali.

Methods: In this study 192 (28.4%) advanced and 256 (37.8%) moderate periodontitis patients were investigated for socio-demographic variables, city of precedence, periodontal parameters, smoking, red complex periodontopathic bacteria, serum antibodies against *Porphyromonas gingivalis* and *Aggregatibacter actinomycetemcomitans* and blood lipids including total cholesterol, HDL, LDL and triglycerides (TG). Those parameters were compared to 229 (33.8%) controls having periodontal health or gingivitis.

Results: Advanced periodontitis had worst periodontal indexes, than moderate periodontitis and controls. Interestingly, higher HDL and TG levels were present in periodontitis. BMI > 30 and smoking were associated with increased HDL, HDL-35, LDL and TG, while glycemia >100 mg/dL associated with HDL, HDL-35 and TG. *Tannerella forsythia* showed a significant association with HDL-35 in bivariate analysis and serum IgG1 against *P. gingivalis* associated with HDL-35 and serum IgG1 against *T. forsythia* associated with TG and serum IgG2 against *A. actinomycetemcomitans* correlated with levels of HDL y HDL-35. In logistic regression the periodontitis patients from Cali presented reduced HDL levels as compared to Bogotá and Medellín patients. Presence of IgG1 antibodies against *P. gingivalis* and *A. actinomycetemcomitans* correlated with reduced HDL levels.

Conclusion: This study confirmed that untreated periodontitis generates alteration in serum lipid levels and systemic bacterial exposure against important periodontopathic bacteria could be the biological link.

Resumen

Objetivo: La periodontitis no tratada parece causar inflamación sistémica, así como alteración de los niveles sanguíneos de lípidos, lo que conduce a un mayor riesgo de enfermedades cardiovasculares. Para empezar a probar esta hipótesis en pacientes Colombianos, se realizó un estudio multicéntrico que incluyó las tres capitales principales: Bogotá, Medellín y Cali.

Métodos: Se estudiaron 192 pacientes con periodontitis crónica avanzada (28.4%) y 256 (37.8%) con periodontitis moderada y en estos se determinaron las variables sociodemográficas, ciudad de procedencia, parámetros periodontales, fumar, presencia de bacterias periodontopáticas, anticuerpos séricos contra *Porphyromonas gingivalis* y *Aggregatibacter actinomycetemcomitans*, así como niveles de lípidos en sangre incluyendo colesterol total, HDL, LDL y triglicéridos (TG). Estos parámetros se compararon con 229 (33.8%) pacientes controles sanos/gingivitis.

Resultados: Los pacientes con periodontitis avanzada tuvieron peores índices periodontales que los de periodontitis moderada y los controles. Mayores niveles de HDL y TG estuvieron presentes en pacientes con periodontitis. El índice de masa corporal >30 y el hábito de fumar se asociaron con aumento de HDL, HDL-35, LDL y TG, mientras la glicemia >100 mg/dL se asoció con HDL, HDL-35 y TG. En el análisis bivariable *Tannerella forsythia* mostró asociación significativa con HDL-35 e IgG1 sérica contra *P. gingivalis* estuvo asociada a HDL-35 así como IgG1 contra *T. forsythia* con TG y la IgG2 contra *A. actinomycetemcomitans* se correlacionó con los niveles de HDL y HDL-35. En la regresión logística se observó que la región de Cali tuvo niveles menores de HDL en comparación con los pacientes de Bogotá y Medellín. La presencia de anticuerpos IgG1 contra *P. gingivalis* y *A. actinomycetemcomitans* se asoció con niveles reducidos de HDL.

Conclusión: Este estudio confirmó que la periodontitis no tratada genera alteración en los niveles de lípidos séricos y la exposición bacteriana sistémica a las bacterias periodontopáticas podría ser el vínculo biológico.

*Corresponding author:

E-mail address: inv-odont@univalle.edu.co (Jaramillo A), institutouibo@gmail.com (Lafaurie G), linamillan@gmail.com (Millán LV), martinardila@gmail.com (Ardila C), aduqued@ces.edu.co (Duque A), novoacamil@hotmail.com (Novoa C), dilote@hotmail.com (López D), adolfoco@yahoo.com (Contreras A).

Introduction

Periodontitis is a chronic infection characterized by an exaggerated gingival inflammatory response to pathogen microbiota, which results in the loss of dental support tissue and, eventually, in the loss of teeth¹; It is associated to other systemic chronic conditions like atherosclerotic cardiovascular disease through common pathophysiological pathways, hence, it may be considered that by improving periodontal health local and systemic inflammation is reduced and, thus, cardiovascular risk² may be reduced. In Colombia, according to the results from the most recent National Study on Oral Health in 1999 (ENSAB III), it was found that in the adult population 50.2% of the people present loss of periodontal insertion and this prevalence increases with age³.

Also, cardiovascular diseases are the first causes of death globally, and the WHO estimates that close to 17.3 million people died due to this cause in 2008, which represents 30% of all deaths worldwide⁴.

The foundation of the association between periodontal disease and other systemic inflammatory conditions is chronic inflammation, given that evidence exists that individuals with periodontitis have greater risk of presenting endothelial dysfunction and cardiovascular diseases. The pathogeny of destructive periodontal disease and atherosclerotic disease, hence, can be related through common inflammatory cascades⁵. Among the inflammatory mediators associated to the risk of cardiovascular events we find serum amyloid A, sICAM-1, IL-6, homocysteine, total cholesterol, LDL cholesterol, and C-reactive protein^{6,7}, which increase in patients with periodontitis inducing pro-coagulation and alterations in lipid metabolism, which can increase risks of cardiovascular events.

The relationship between periodontitis and dyslipidemia seems to be two-way relation, that is, it is not clear if periodontal disease affects lipid metabolism or if the conditions associated to dyslipidemia damage the dental support tissue⁸.

Although it has been suggested that alterations in lipid metabolism and periodontitis may be associated through common physiopathological mechanisms, which explains increased risk of cardiovascular disease in patients with periodontitis^{9,10}, no published evidence exists on the association between periodontitis and dyslipidemia in Colombia. Due to these reasons, this research sought to study the association between dyslipidemia and untreated periodontitis from a sample of the Colombian population.

Materials and Methods

An observational study was carried out with a sample of 677 patients who attended the dental clinics at Universidad El Bosque and Pontificia Universidad Javeriana in Bogotá, Universidad CES and Universidad de Antioquia in Medellín, and Universidad del Valle in Cali, during the period comprised between January 2009 and March 2012.

The sample size was calculated with a 5% alpha error and 80% beta error to find associations between periodontal risk factors and dyslipidemia with an expected alteration probability $\geq 15\%$ in lipid profile parameters evaluated in the control population, for an OR ≥ 2 .

Selection criteria: The study included the subjects who accepted their voluntary participation and who attended the dental clinics at the participating universities. Inclusion criteria for patients with periodontitis included having any degree of severity from moderate to severe and at least 14 teeth present in the mouth. The group of control patients (healthy or with gingivitis) was constituted by patients who had maximum four sites with 4 mm pockets in all the sites examined. The study excluded individuals who had received periodontal treatment or systemic antibiotics six months prior to the periodontal exam, and pregnant women or those individuals presenting systemic diseases like HIV or AIDS, cancer, risk of infectious endocarditis and autoimmune diseases.

Survey to collect information: Via a structured survey, previously validated through a pilot study, We obtained data corresponding to variables of age, geographic region, gender, socioeconomic level, and smoking habits.

Periodontal clinical evaluation: The clinical exam was performed by a periodontist in each of the participating universities, who was trained and calibrated before the start of the study. The exam was conducted of the full mouth, in six sites per tooth, using a periodontal probe UNC 15. The indices obtained were the Loe & Silness gingival index (GI), the plaque index (PI), pocket depth (PB), clinical insertion level (CIL), and bleeding on probing (BOP).

According to the disease severity, patients were distributed into two categories according to the average of the insertion level of the affected sites: slight to moderate chronic periodontitis and severe periodontitis. The control group was made up of patients diagnosed as healthy or with gingivitis.

Evaluation of periodontal infection indicators

Sample of subgingival plaque : After the clinical evaluation, the six deepest subgingival sites (pockets > 5 mm) were taken in patients with periodontitis and one site per sextant healthy patients; the supragingival plaque was eliminated with a sterile curette and the sample taking site was isolated. Sterile absorbing paper tips were inserted during 20 seconds; these were collected in a sterile Eppendorf tube for their processing in two microbiology laboratories using standardized Polymerase Chain Reaction (PCR) methods.

Polymerase Chain Reaction: Polymerase Chain Reaction was carried out according to the protocol by Ashimoto *et al.*, in 1996. with a final reaction volume of 25 μL of which 5 μL corresponded to the sample and 20 μL to the reaction mixture. For *Porphyromonas gingivalis*, *Tannerella forsythia*, and *Treponema denticola*, the reaction mix was composed of buffer PCR 1X u50 mMgCL, 10 mM Tris-HCl (pH 9.0 at

25 °C), 1.5 mM MgCl₂ and 0.1% Triton (R) X-100, 0.25 U of Taq DNA polymerase, 1.5 mM of MgCl₂, 0.2 mM from each deoxyribonucleotide and 2 μM from each primer. For *Aggregatibacter actinomycetemcomitans*, the reaction mix was composed of buffer PCR 1X (50 mM KCl, 10 mM Tris-HCl (pH 9.0 at 25 °C), 1.5 mM MgCl₂ and 0.1% Triton (R) X-100), 0.25 U of Taq DNA polymerase, 2.25 mM of MgCl₂, 0.2 mM of each deoxyribonucleotide and 2 μM of each primer.

Sample amplification was carried out in a thermocycler (MyCycler Thermal Cycler, Bio-Rad). The temperature cycles for *P. gingivalis*, *T. forsythia*, and *T. denticola* included an initial denaturing step at 95° C for 2 min, followed by 36 denaturing cycles at 95° C for 30 seconds, alignment at 60° C for 1 min, extension at 72° C for 1 min and a final passage at 72° C for 2 min. The temperature for *A. actinomycetemcomitans* included an initial passage at 95° C for 2 min, followed by 36 cycles at 94° C for 30 seconds, 55° C for 1 min, 72° C for 2 min, and 72° C for 10 min.

The PCR products were evaluated via electrophoresis in agarose gel at 1.5% in TAE buffer (Tris-acetate EDTA) stained with 0.5 mg/mL of ethidium bromide and were visualized via trans-illumination with 300 nm UV light. As positive control, we employed DNA from reference strains ATCC 33277 from *P. gingivalis*, ATCC 43037 from *T. forsythia*, ATCC 29522 from *A. actinomycetemcomitans*, and ATCC 35405 from *T. denticola*. As negative control, we employed sterile bi-distilled water. The finding of *P. gingivalis* in the samples was evidenced by the presence of a band corresponding to 404 bp, *T. forsythia* at 641 bp, *T. denticola* at 316 bp, and *A. actinomycetemcomitans* at 557 bp, compared to a marker with 1-Kb molecular weight.

Antibody levels against periodontal pathogens: Levels of serum antibodies were conducted at the reference centers, where these methods were previously standardized to assess levels of serum antibodies for the four microorganisms studied. The technique used was indirect ELISA, covering 96-well plates (Immulux Dynex®) with *P. gingivalis*, *T. forsythia*, *T. denticola*, and *A. actinomycetemcomitans* homogenized at 2 μg/mL concentration incubated overnight at 4 C. The specific bonding sites were blocked with 200 μL of albumin at 1% in PBS-Tween (phosphate salt buffer, pH 7.4; Tween 0.05%) at 37 °C for 2 hrs.

Sera from patients were diluted in blocking solution (1:1,000) and incubated at 37 °C for 1 hr. Thereafter, the plates were incubated with diluted biotinylated 1:1,000 antihuman IgG1 antibody or antihuman IgG2 (Sigma®) at 37° C for 1 hr. Diluted 1:1,000 streptavidin-peroxidase conjugate (Vector®) was added and incubated at room temperature for 1 hr.

The plates were developed with OPD substrate (O-Phenylenediamine) for 10 min and the reaction was halted with H₂SO₄ 2.5 M, to proceed to its reading at 490 nm in a Stat Fax 2100 ELISA reader. To determine IgG1 and IgG2 levels in the samples, specific for each of the microorganisms, standard reference sera were used presenting high levels of IgG1 or IgG2, at different dilutions whose optical density

values were plotted on a linear regression curve.

Levels of IgG subtypes in the samples were calculated from the formula $Y = M * X + C$, obtained with two standard reference sera, and reported as concentration in E.U./mL (ELISA Units). The specific bonds of the assay were determined by conducting ELISA without serum sample and replacing it with water, to subtract this value for each of the serum samples analyzed, and each sample was analyzed in triplicate.

Evaluation of lipid profile: Blood samples were taken from each patient via venipuncture, and serum lipid profile was analyzed including total cholesterol, LDL, HDL, and triglycerides (TG). Tests were conducted in the same national level reference laboratory.

Statistical analysis: A database was created on ACCESS® 2007, which was evaluated periodically using the SAS® statistical package version 9.1. Then, the database was imported to the Stata® program version 11.2, to perform the descriptive, bivariate, and multivariate analysis.

Logistic regression models were developed to establish the association of the independent variables with the presence and severity of periodontitis (severity based on insertion level), as well as with the level of inflammatory activity in relationship with pocket depth, level of bleeding, and periodontal infection markers adjusted for age, gender, income level, and geographic region.

Serum was evaluated for levels of total cholesterol, TG, HDL, and LDL among the different groups of healthy/gingivitis patients, with moderate periodontitis and with severe periodontitis adjusting for confounding factors. The cutoff points to consider lipid levels as risk factor were for HDL cholesterol for cardiovascular risk >40 mg/dL in women and >50 mg/dL in men and for high cardiovascular risk HDL-35 > 35 mg/dL. For triglycerides, the cutoff point was 150 mg/dL, for LDL cholesterol <130 mg/dL, and for total cholesterol <200 mg/dL.

Results

A total of 448 patients with periodontitis were included of which 192 (28.4%) had advanced periodontitis and 256 (37.8%) moderate periodontitis. Risk factors taken in consideration were socio-demographics, periodontal parameters, smoking, presence of periodontopathic organisms, and serum antibodies against these pathogens, as well as systemic inflammation factors. These groups of patients were compared between and against 229 (33.8%) periodontally healthy or gingivitis controls.

Medellín city contributed the highest number of cases of advanced periodontitis with 88 cases (45.8%), followed by Bogotá with 73 (38%), while Cali only had 31 cases (16.1%). Patients with advanced and moderate periodontitis were on the average older (48 years) than the control patients (45.5 yrs); this difference was significant (Table 1) with females predominating.

Table 1. Socio-demographic variables of study subjects

Parameter	Control group		Periodontitis			
	Total	%	Moderate		Advanced	
			Total	%	Total	%
Subjects	229/677	33.8 ^b	256/677	37.8 ^{a,c}	192/677	28.4 ^b
Region:						
Medellín	51/229	22.3 ^{b,c}	86/256	33.6 ^a	88/192	45.8 ^a
Bogotá	134/229	58.5	102/256	39.8	73/192	38,00
Cali	44/229	19.2	68/256	26.6	31/192	16.1
Gender						
Female	163/229	71.2	168/256	65.6	105/192	54.7
Male	66/229	28.8	88/256	34.4	87/192	45.3
Socioeconomic level						
B	139/229	60.7	189/256	73.8	154/192	80.2
M	90/229	39.3	67/256	26.2	38/192	19.8
Age (Mean±SD)	45.50±8.02 ^{b,c}		48.2±9.33 ^a		48.68±9.93 ^a	

p < 0.05 statistical test ANOVA, Kruskal Wallis Mann Whitney U or Chi squared, a. Differences with Control, b. Differences with Moderate periodontitis c. Differences with Advanced periodontitis

Table 2 evidences that control patients had better periodontal clinical parameters than patients with moderate and advanced periodontitis, given that significant differences were found in average pocket depth, clinical insertion level, bleeding on probing, and in gingival index. Loss of clinical insertion was of 5 ± 1.1 mm in advanced periodontitis, 2.8 ± 0.6 mm in moderate, and 1.3 ± 0.6 mm in the control group.

Table 3 presents the bivariate analysis correlating socio-demographic variables, periodontal, systemic, environmental and microbiological factors against lipid profiles in these patients. Statistically significant differences were noted among regions for total cholesterol, HDL, HDL 35, and TG. Additionally, an association was found among age and total cholesterol and LDL values. The periodontal clinical variables, as a set, were associated to cholesterol and LDL levels; however, those differences were not associated to the periodontal clinical state given that controls revealed higher levels of total cholesterol and LDL than patients with periodontitis. The BMI >30 is associated to cholesterol, HDL, HDL-35, LDL, and TG, while glycemia >100 mg/dL is associated to HDL, HDL-35, and TG. Finally, smoking is associated to levels of HDL, HDL-35, and LDL for increased cardiovascular risk.

Of the microorganisms studied, only *T. forsythia* had

Table 2. Clinical parameters according to periodontal diagnosis

Parameter/Diagnosis	Control group	Periodontitis	
		Moderate	Advanced
No. of teeth	25.4±3.6	24.3±4.1	23.2±3.8
Pocket depth (Mean±SD)	2.1±0.4 ^{b,c}	2.8±0.5 ^{a,c}	4.1±0.9 ^{a,b}
Clinical insertion level (Mean±SD)	1.3±0.6 ^{b,c}	2.8±0.6 ^{a,c}	5.0±1.1 ^{a,b}
Bleeding on probing (% positive)	33±25 ^{b,c}	56±26 ^{a,c}	70±28 ^{a,b}
Gingival index (% positive)	39±30 ^{b,c}	61±29 ^a	68±34 ^a

p < 0.05 statistical test ANOVA, Kruskal Wallis, Mann Whitney U or Chi squared.
 p < 0.05 statistical test ANOVA, Kruskal Wallis, Mann Whitney U or Chi squared. a. Differences with Control, b. Differences with Moderate periodontitis c. Differences with Advanced periodontitis

a significant association with HDL-35 in the bivariate analysis. With respect to IgG1 serum against *P. gingivalis*, it is associated to HDL-35 and IgG1 serum against *T. forsythia* is associated to TG (Table 4). Additionally, the presence of IgG2 antibodies against *A. actinomycetemcomitans* was correlated to high levels of HDL and HDL-35.

Table 4 presents the risk factors adjusted in the logistic regression. The region of Cali was constituted as a protective factor for HDL levels of medium cardiovascular risk. Having high levels of IgG1 antibodies against *P. gingivalis* and medium and high levels for *A. actinomycetemcomitans* increases the risk of having low HDL levels. Regarding HDL levels that indicate high cardiovascular risk, the associated variables were overweight, glycemia >100 mg/dL, IgG2 for *A. actinomycetemcomitans*, and smoking.

Discussion

Periodontitis has been associated to hypercholesterolemia and hypertriglyceridemia, however these effects on lipid metabolism raise controversy. Thus, further laboratory research is required along with intervention clinical studies treating periodontal diseases to reveal the exact mechanisms that relate periodontitis with dyslipidemia and atherosclerosis. Evidence until now suggests that oral hygiene and periodontal diseases are associated to alterations in lipid metabolism. For example, a study with adult subjects participating on cohort in Japan found that those self-reporting better tooth-brushing habits had lower levels of triglycerides¹¹.

Our observational study revealed that untreated periodontitis is associated to the alteration of important lipid markers related to cardiovascular disease. Patients with periodontitis have lower levels of high-density lipoprotein (HDL) or anti-atherogenic lipoprotein; however, this is not associated to the unexpected increase of low-density lipoprotein (LDL) or pro-atherogenic lipoprotein. (table 3) Other researchers have found association between alterations in lipid metabolism and periodontitis. In a population-based study

Table 3. Bivariate analysis of socio-demographic factors, periodontal state, systemic, environmental factors.

Variable	ChL	HDL	HDL-35	LDL	TG
Socio-demographic					
Region	<0.10*	<0.05**	<0.05**	NS	<0.05**
Age	<0.0001†	NS	NS	<0.0001†	NS
Gender	NS	<0.05**	<0.0001†	NS	<0.0001†
Socioeconomic level	NS	<0.05**	NS	NS	NS
Periodontal state					
Diagnosis	NS	NS	NS	<0.05**	NS
NI Extension	<0.10*	NS	NS	<0.05**	NS
Percentage of sites with	<0.05**	NS	NS	<0.05**	NS
Average pocket depth	<0.05**	NS	NS	<0.10*	NS
IG Percentage	<0.10*	NS	NS	NS	NS
Hemorrhage percentage	<0.10*	NS	NS	<0.10*	NS
Systemic factors					
BMI	<0.05**	<0.05**	<0.05**	<0.05**	<0.0001†
Glycemia	NS	<0.05**	<0.05**	NS	<0.0001†
Antihypertensive	<0.10*	NS	NS	NS	NS
Cholesterol Medications	NS	<0.05**	NS	NS	NS
Environmental factors					
Smoking	NS	<0.05**	<0.10*	<0.05**	NS
Bacterial factors					
<i>P. gingivalis</i>	NS	NS	NS	NS	NS
<i>T. forsythia</i>	NS	NS	<0.05**	NS	NS
<i>T. denticola</i>	NS	NS	NS	NS	NS
<i>A. actinomycetemcomitans</i>	NS	NS	NS	NS	NS
Serum antibodies					
IgG1 <i>P. gingivalis</i>	NS	<0.05**	NS	NS	NS
IgG1 <i>T. forsythia</i>	NS	NS	NS	NS	<0.05**
IgG1A.	NS	NS	NS	NS	NS
IgG2 <i>P. gingivalis</i>	NS	NS	<0.05**	NS	NS
IgG2 <i>T. forsythia</i>	NS	NS	NS	NS	NS
IgG1A.	NS	<0.05**	<0.10*	NS	NS

Bacterial factors and serum antibodies as indicators of dyslipidemia. ChL = Total cholesterol; HDL = HDL cholesterol <40 mg/dL in women, <50 mg/dL in men; HDL-35 = HDL cholesterol <35 mg/dL; LDL = LDL cholesterol; TG = Triglycerides, * p < 0.10 ** p < 0.05 † p < 0.0001.

in Korea, the authors reported adjusted ORs of 1.38 (95%CI: 1.17–1.62) for hypertriglyceridemia and 1.34 (95%CI: 1.14–1.56) for low HDL cholesterol¹². Also, the United States, an adjusted OR was found between moderate periodontitis and low levels of HDL at 1.42². In Brazil, in a study of cases and controls no associations were found between the severity of the periodontitis and the lipid serum levels, possibly due to the limited size and the procedure to select the of subjects¹³.

The HDL lipoprotein is considered anti-atherogenic because it neutralizes LPS in the circulation¹⁴ and prevents LDL oxidation¹⁵, as well as antagonizes cholesterol transport¹⁶, because it accepts cholesterol from the cell membrane during its elimination. This process is facilitated by passive diffusion of cholesterol toward HDL and actively by the interaction of lipoproteins poor in apolipoprotein A1 (ApoA1), preBeta-HDL or ABCA1, which facilitates cholesterol removal¹⁷. HDL cholesterol is esterified in the blood circulation and it is directly transported to the liver via LDL for excretion. Thus, HDL promotes elimination of cholesterol and a failure in this elimination route might relate to thickening of the vascular wall and to the appearance of early atherosclerosis-type lesions¹⁸.

During chronic inflammation, as in the case of untreated periodontitis, changes occur in lipoprotein distribution and in the proportions of cholesterol subtypes¹⁹. Infection increases HDL catabolism and can reduce levels of HDL cholesterol²⁰. Thus, the main protein, ApoA1, is displaced by an increase in lipid serum amyloid A (SAA), which is synthesized in big proportions in response to an increase of pro-inflammatory proteins²¹. Triglycerides also increase

when there is inflammation and infection, phenomena in which cholesterol- and HDL-rich complexes are formed, and are substrate for hepatic lipase that when activated stimulates the formation of poor lipids that suffer accelerated catabolism through the kidney²². Within this scenario, the infection and inflammation cause dramatic changes in HDL levels and in its metabolism²³. Infection also induces some other atherogenic changes in lipoprotein profiles and these can be some of the mechanisms that link chronic inflammation to the development of atherosclerosis. Various pathogens are capable of causing alterations in lipid metabolism, among them are *Chlamydia pneumoniae*, *Helicobacter pylori*²⁴, and periodontal pathogens *P. gingivalis*, *T. forsythia*, and *A. actinomycetemcomitans* that have been found associated to diverse lipid metabolites^{25,26}. With respect to the levels of antibodies to the periodontal pathogens, in the present study, *T. forsythia* had a significant association with HDL-35 in the bivariate analysis (Table 3), while Serum IgG1 against *P. gingivalis* was associated to HDL-35 and Serum IgG1 against *T. forsythia* was associated to TG (Table 3). Additionally, the presence of IgG2 antibodies against *A. actinomycetemcomitans* was correlated to high levels of HDL and HDL-35 as shown in Table 4 the logistic regression analysis.

Given the temporality of the present study, it may not be ignored that alterations in lipid metabolism occur due to a cause different from periodontitis, but after the adjustment these variables are associated on the regression model.

In conclusion, this study confirmed that untreated periodontitis is associated to alterations of lipid metabolism; additionally, it was shown that bacterial systemic exposure exists to periodontopathic microorganisms, as revealed by the levels of IgG antibodies against *P. gingivalis* and *A.*

Table 4. Logistic regression. Risk factors for HDL cholesterol

HDL cholesterol. Medium risk		Adjusted OR	[95% CI]
VARIABLE	Reference		
Region	Bogotá		
Medellín		0.95**	0.66–1.36
Cali		0.54**	0.35–0.84
Antibodies			
IgG1 <i>P. gingivalis</i>	Low		
High		2.62**	1.39-4.93
IgG2 <i>A. actinomycetemcomitans</i>	Low		
Medium		1.97**	1.24-3.13
High		2.09**	1.12-3.91
HDL-35- High-risk cholesterol			
ÍMC	Normal		
Overweight> 25 < 30		1.58**	0.98-2.53
Glycemia	70-100 mg/dl		
>100 mg/dl		2.01**	1.19-3.41
Antibodies			
IgG2 <i>A. actinomycetemcomitans</i>	Low		
Medium		1.77**	1.01-3.08
Smoking			
Positive	Negative		
		1.84**	1.11-3.04

*p < 0.10 ** p < 0.05 † p < 0.0001

actinomycetemcomitans associated to possibly increased cardiovascular risk in the regression model. More research is required on this subject to confirm the hypothesis that untreated periodontitis alters lipid metabolism via infection and inflammation.

Conflict of interest:

The authors declare that there is no real or potential conflict of interest regarding the possible publication of this work.

Acknowledgments

This research was sponsored by COLCIENCIAS, by the project: Risk factors in chronic periodontitis in Colombia (againststct 422-2008), by the project: genotyping of *FimA* of *p. givialis* (CI16a5) and cardiovascular risk markers in periodontitis – Universidad del Valle (CI 1688), and by personnel contributions in the participating universities.

References

1. Van Dyke TE, Serhan CN. Resolution of inflammation: a new paradigm for the pathogenesis of periodontal diseases. *J Dent Res.* 2003; 82(2): 82-90.
2. Aiuto, F. D', Sabbah W, Netuveli G, Donos N, Hingorani, A. D., Deanfield J, *et al.* Association of the Metabolic Syndrome with Severe Periodontitis in a Large U.S. Population-Based Survey. *J Clin Endocrinol Metab.* 2008; 93(10): 3989-94.
3. Ministerio de Salud de Colombia. III Estudio Nacional de Salud Bucal - ENSAB III. II Estudio Nacional de Factores de Riesgo de Enfermedades Crónicas -ENFREC II: Prevalencia de Diabetes Mellitus y de glucosa alterada en ayunas T. V. 3 ed. Bogotá: Ministerio de Salud de Colombia- Centro Nacional de Consultoría; Bogotá, 1999.
4. Who . Cardiovascular diseases (CVDs). Geneva: World Health Organization; 2013.
5. Nesbitt MJ, Reynolds MA, Shiao H, Choe K, Simonsick EM, Ferrucci L. Association of periodontitis and metabolic syndrome in the Baltimore Longitudinal Study of Aging. *Aging Clin Exp Res.* 2010; 22(3): 238-42.
6. Noack B, Genco RJ, Trevisan M, Grossi S, Zambon JJ, De Nardin E. Periodontal infections contribute to elevated systemic C-reactive protein level. *J Periodontol.* 2001; 72(9): 1221-7.
7. Ridker PM, Hennekens CH, Buring JE, Rifai N. C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women. *N Engl J Med.* 2000; 342(12): 836-43.
8. Izumi Y, Nagasawa T, Umeda M, Kobayashi H, Takeuchi Y, Yashiro R, *et al.* Periodontitis and cardiovascular diseases: The link and relevant mechanisms. *Jpn Dent Sci Rev.* 2009; 45(2): 98-108.
9. Lösche W, Karapetow F, Pohl A, Pohl C, Kocher T. Plasma lipid and blood glucose levels in patients with destructive periodontal disease. *J Clin Periodontol.* 2000; 27(8): 537-41.

10. Katz J, Flugelman MY, Goldberg A, Heft M. Association between periodontal pockets and elevated cholesterol and low density lipoprotein cholesterol levels. *J Periodontol.* 2000; 73(5): 494-500.
11. Kobayashi Y, Niu K, Guan L, Momma H, Guo H, Cui Y. Oral health behavior and metabolic syndrome and its components in adults. *J Dent Res.* 2000; 91(5): 479-84.
12. Kwon YE, Ha JE, Paik DI, Jin BH, Bae KH. The relationship between periodontitis and metabolic syndrome among a Korean nationally representative sample of adults. *J. Clin. Periodontol.* 2011; 38(9): 781-6.
13. Machado AC, Quirino MR, Nascimento LF. Relation between chronic periodontal disease and plasmatic levels of triglycerides, total cholesterol and fractions. *Braz Oral Res.* 2005; 19(4): 284-9.
14. Levine DM, Parker TS, Donnelly TM, Walsh A, Rubin AL. In vivo protection against endotoxin by plasma high density lipoprotein. *In vivo* protection against endotoxin by plasma high density lipoprotein. *PNAS.* 1993; 90(24): 12040-4.
15. Mackness MI, Durrington PN, Mackness B. How high-density lipoprotein protects against the effects of lipid peroxidation. *Curr Opin Lipidol.* 2000; 11(4): 383-8.
16. Fielding CJ, Fielding PE. Molecular physiology of reverse cholesterol transport. *J Lipid Res.* 1995; 36(2): 211-28.
17. Mendez AJ. Cholesterol efflux mediated by apolipoproteins is an active cellular process distinct from efflux mediated by passive diffusion. *J Lipid Res.* 1997; 38(9): 1807-21.
18. van Dam MJ, de Groot E, Clee SM, Hovingh, G Kees, Roelants R, *et al.* Association between increased arterial-wall thickness and impairment in ABCA1-driven cholesterol efflux: an observational study. *Lancet.* 2002; 359(9300): 37-42.
19. Sammalkorpi K, Vhignen V, Kerttula Y, Nikkilä E, Taskinen MR. Changes in serum lipoprotein pattern induced by acute infections. *Metabolism.* 1988; 37(9): 859-865.
20. Feingold KR, Krauss RM, Pang M, Doerrler W, Jensen P, Grunfeld C. The hypertriglyceridemia of acquired immunodeficiency syndrome is associated with an increased prevalence of low density lipoprotein subclass pattern B.. *J Clin Endocrinol Metab.* 1993; 76(6): 1423-7.
21. Betts JC, Lukey PT, Robb LC, McAdam RA, Duncan K. Evaluation of a nutrient starvation model of Mycobacterium tuberculosis persistence by gene and protein expression profiling. *Mol Microbiol.* 2002; 43(3): 717-31.
22. Newnham HH, Barter PJ. Synergistic effects of lipid transfers and hepatic lipase in the formation of very small high-density lipoproteins during incubation of human plasma. *Biochim. Biophys. Acta.* 1990; 1044(1): 57-64.

23. Ard JD, Rosati R, Oddone EZ. Culturally-sensitive weight loss program produces significant reduction in weight, blood pressure, and cholesterol in eight weeks. *J Natl Med Assoc.* 2000; 92(11): 515-23.
24. Laurila A, Bloigu A, Näyhä S, Hassi J, Leinonen M, Saikku P. *Chlamydia pneumoniae* and *Helicobacter pylori* infections in Sámi and Finnish reindeer herders. *Int J Circumpolar Health.* 1997; 56(3): 70-5.
25. Beck JD, Offenbacher S. Relationships among clinical measures of periodontal disease and their associations with systemic markers.. *Ann Periodontol.* 2002; 7(1): 79-89.
26. Pussinen PJ, Jauhiainen M, Vilkuna-Rautiainen T, Sundvall J, Vesanen M, Mattila K, et al. Periodontitis decreases the antiatherogenic potency of high density lipoprotein. *J Lipid Res.* 2003; 45(1): 139-47.
-